Gonghao Lu,^a* Akira Katoh,^b Zhiqiang Zhang,^c Zhizhi Hu,^c Peng Lei,^c and Masaru Kimura^c

^aOrganic Nanotube Team (ONT), Nanoarchitectonics Research Center (NARC), National Institute of Advanced Industrial Science and Technology (AIST), Tsukuba Central 5-2, 1-1-1 Higashi, Tsukuba, Ibaraki 305-8565, Japan
^bDepartment of Materials and Life Science, Faculty of Science and Technology, Seikei University,

3-3-1 Kichijoji-kitamachi Musashinno, Tokyo 180-8633, Japan

^cDepartment of Applied Chemistry, University of Science and Technology Liaoning,

Anshan 114044, People's Republic of China

*E-mail: ghlu-ro@aist.go.jp

Received July 1, 2009

DOI 10.1002/jhet.385

Published online 18 June 2010 in Wiley InterScience (www.interscience.wiley.com).



Lophine hydroperoxides underwent base-triggered 1,5-phenyl migration in DMSO to afford imidazolones in high yields, instead of amidines with chemiluminescence (CL). The corresponding imidazolols were believed to intermediates and they were successfully obtained by treating the peroxides with DMSO without the base. The diminished CL was because of the reduction of the hydroperoxides with DMSO. The imidazolols subsequently underwent smooth base-mediated rearrangement to afford imidazolones. Furthermore, the chiral imidazolols provided stereoselective imidazolones in high enantiomeric excess (>92%), which supported the mechanism of an intramolecular ring for the migration.

J. Heterocyclic Chem., 47, 932 (2010).

INTRODUCTION

The chemiluminescent reactions of lophine hydroperoxides (1) have attracted considerable attention for many decades [1,2]. Many studies have been reported [3] and these studies mainly focused on the chemiluminescence (CL) efficiency. The other reactions occurring in this chemiluminescent process are still not clear. Recently, we reported that 1 underwent three different but simultaneous reactions upon treatment with a base to yield the corresponding amidines (2) accompanied with CL, imidazoles (3) with singlet oxygen, and a trace of imidazolones (4) (Scheme 1) [4]. However, the CL of peroxides 1 can be observed by base-triggered reactions in typical solvents, but not in DMSO. It is well known that a CL system like dioxetane can provide the most efficient CL in DMSO [5]. We first believed that a polar aprotic solvent like DMSO should enhance the CL efficiency of lophine peroxides. However, the CL efficiency of hydroperoxides 1 in was so low that it could not be detected. The results were quite unexpected, and therefore, they attracted our attention. The subsequent investigation showed that hydroperoxides 1 underwent an exclusive 1,5-phenyl migration in DMSO to afford imidazolones 4 in high yields via imidazolols (5) as intermediates (Scheme 2) [6]. Further investigation showed that these imidazolols 5 could be easily obtained under milder conditions, when treated with DMSO without the trigger base [6b]. In addition, these imidazolols 5 subsequently underwent base-mediated rearrangement to afford imidazolones in high yields.





Although imidazolol has been long proposed as one of the intermediate byproducts in the CL reaction of lophine, it has not been obtained thus far because it is unstable under basic reaction conditions [1b]. The present research provides an easy method to obtain these intermediates, and this should help in completely understanding the reactions occurring in the CL reaction of lophine peroxides. In addition, the sigmatropic migration of an aryl group is known for 1,2,3,4,5-pentaphenylcyclo-pentadienol [7], which is a π -conjugated analogue of 5; however, its stereochemistry has not yet been investigated. In a previous investigation, [6a] the 1,5-phenyl rearrangement of silyl-protected hydroperoxides was reported. However, the stereospecificities were low under thermal conditions and the enantiomeric excesses (EEs) were less than 60%; these factors made it difficult to understand the stereochemistry. In the previous study, we first believed that the low stereospecificities resulted from the racemization of imidazolones through a ring opening/closing sequence, and therefore, a control experiment was carried out with imidazolone; however, it was found that it did not racemize. Therefore, we believed that the alkoxide racemized through a ring opening/closing sequence under strong thermal conditions that lowered the stereochemical specificities. In this study, the absence of strong thermal activation prevents ring opening from occurring, as a result of which imidazolols can serve as a good system for the study of 1,5-phenyl migration.

Furthermore, it should be noted that many natural compounds include imidazolol and/or imidazolone moieties [8]. In this article, we report the preparation and the stereoselective 1,5-phenyl rearrangement of imidazolols. We expect that imidazolols will find numerous applications in the synthesis of natural compounds.

RESULTS AND DISCUSSION

The reaction of the peroxides 1 proceeded smoothly proceeded to afford imidazolols in good yields in DMSO at room temperature (entries 1–8, Table 1). The reaction completed within 4–6 h. Aromatic substituents containing electron-donating as well as electron-withdrawing groups underwent the elimination of oxygen facilely. In the case of unstable 1c, which is difficult to isolate, [2] 5c was consequently prepared by treating the mixture by the photoxidation of the corresponding imidazole with DMSO. In the cases of 1g, h, the mixtures with their isomers (1'g, h) were used to afford mixtures of 5g, h with their isomers (5'g, h), as shown in Scheme 3.

On treatment with the base, the imidazolols **5** were smoothly converted into the imidazolones **4** via phenyl migration from C4 to C5 within 3 h in good yields in DMSO at room temperature (Table 1). From the Woodward–Hoffmann rules, the phenyl migration is recognized as a thermally allowed 1,5-sigmatropic migration.

To elucidate the stereochemistry of the migration, imidazolols **5g**, **h** were successfully isolated from their mixtures with their isomers **5'g**, **h** by crystallization from 2-propanol, respectively. The chiral imidazolols (**5g** and **h**) were isolated using a chiral HPLC column. To determine the absolute configurations, the circular dichroism (CD) spectra were recorded, and the calculated CD spectra were processed using Gaussian 03w software package [6a,9]. The absolute configurations were assigned by comparing the experimental CD spectra with the calculated ones (for details, see Experimental section). When the chiral imidazolols (**5g** and **h**)

 Table 1

 Preparation of imidazolols and imidazolones.

Entry	Reactants	Product ^a	Yield (%) ^b	
1	1a	5a	77	
2	1b	5b	87	
3	1c	5c	64	
4	1d	5d	76	
5	1e	5e	70	
6	1f	5f	72	
7	1g ^c	5g ^c	78	
8	1h ^c	5h ^c	82	
9	5a	4a	78	
10	5b	4b	85	
11	5c	4c	68	
12	5d	4d	73	
13	5e	4 e	70	
14	5f	4f	76	
15	5g ^c	4g	79	
16	5h ^c	4h	74	

 a The structure of products were determined from spectral data (1 H NMR, MS, and E.A.).

^b Isolated yields after column chromatography.

^c Mixture with its isomer.



were subjected to phenyl migration under the action of the base (Scheme 4), the corresponding imidazolones 4gand **h** were exclusively obtained in >92% EE in DMSO (Table 2). When compared with the previous study in which silyl-protected peroxides under thermal condition were used, the stereoselectivities were largely increased. The reaction mechanism was confirmed to be stereoselective 1,5-phenyl migration via an intramolecular ring (Scheme 5).

In conclusion, we have demonstrated that an imidazolol derivative can easily be prepared in good yield from the corresponding peroxide. The imidazolol can be smoothly converted to an imidazolone via a stereoselective phenyl migration from C4 to C5 under the action of a base.

EXPERIMENTAL

General procedure. All melting points were measured using a Yanagimoto micro melting point apparatus. The IR spectra were recorded by a JASCO FT/IR-5000 spectrophotometer. The UV-vis spectra were measured by a JASCO V-530 spectro-photometer. The ¹H and ¹³C NMR spectra were recorded using a Varian MERCURY (FT, 300 MHz) spectrometer or a Varian VXR-500 (FT, 500 MHz) spectrometer. Elemental analyses were performed by a Perkin Elmer CHNS/O



 Table 2

 The analyses of the enantiomeric excess.

		Products ^{a,b}		
Entry	Imidazolols	R-4	S-4	EE (%)
1	R-5g	2	98	96
2	S-5g	97	3	94
3	R-5h	3	97	94
4	S-5h	96	4	92

^a Yields calculated on the base of HPLC integral quantity.

 $^{\rm b}$ The conversion was estimated to be 100% because of no other peak appearing on the $^{\rm 1}{\rm H}$ NMR spectra.

Analyzer 2400. The fast atom bombardment (FAB) mass spectra were recorded by a Micromass 70-SE. 2,4,5-Triarylphenyl-1*H*-imidazoles were prepared by the method of Davidson *et al.* [6,8a]. 4-Hydroperoxy-2,4,5-triphenyl-4*H*-imidazoles were prepared by the method of White and Harding [2]. HPLC analyses were performed on a Hitachi 655 liquid chromatography and recorded on a Hitachi 561 recorder; Column for EE and optical resolution was Daicel Chiralpak AD-H: 4.6 mm \times 250 mm. CD spectra were recorded on a JASCO J-820 spectropolarimeter.

Preparation of imidazolols. A solution of hydroperoxides **1** (0.2 mmol) in DMSO (5 mL) was stirred for 4–6 h at room temperature. After the reaction, the solution was poured into water, and the crude product **5** was obtained by filtration. The crude product was purified by chromatography (silica, hexane:AcOEt = $10 \sim 8$:1).

2,4,5-Triphenyl-4-hydroxy-4H-isoimidazole (5a). mp 128–130°C; IR (KBr) 1613 (C=N) cm⁻¹, ¹H NMR (500 MHz, CDCl₃) δ 7.16–7.22 (m, 3H), 7.34 (dd, J = 7.5, 5.5 Hz, 2H), 7.38 (t, J = 7.5 Hz, 2H), 7.47 (t, J = 7.5 Hz, 1H), 7.49 (d, J = 7.5 Hz, 2H), 7.56 (t, J = 7.5 Hz, 1H), 8.16 (d, J = 7.5 Hz, 2H), 8.41 (d, J = 7.5 Hz, 2H), 6.49 (br s, 1H); ¹³C NMR (300 MHz, CDCl₃) δ 107.66 (s), 124.9 (d), 128.3 (d), 128.6 (d), 128.7 (d), 128.8 (d), 129.8 (d), 130.2 (d), 130.6 (d), 130.9 (s), 132.5 (d), 133.0 (d), 137.6 (s), 172.5 (s), 194.3 (s); UV-vis λ_{max} (EtOH) 281 (log ϵ 4.31) nm; MS (FAB) *m/z* 313

 (M^++1) ; E.A. Calcd for $C_{21}H_{16}N_2O$: C, C, 80.75; H, 5.16; N, 8.97. Found: C, 80.70; H, 5.14; N, 8.96.

2-(*p*-*Nitrophenyl*)-4-hydroxy-4,5-diphenyl-4H-isoimidazole (5b.) Pale yellow powder, mp 168–169°C; IR (KBr) 1633 (C=N), 1524 (NO₂), 1350 (NO₂) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.33–7.38 (m, 3H), 7,43 (dd, J = 8.4, 2.0 Hz, 2H), 7.56 (t, J = 8.1 Hz, 2H), 7.66 (t, J = 8.1 Hz, 1H), 8.09 (d, J= 9.2 Hz, 2H), 8.19 (d, J = 9.2 Hz, 2H), 8.36 (d, J = 8.1 Hz, 2H), 6.94 (s, 1H); MS (FAB) *m*/*z* 358 (M⁺+1); E.A. Calcd for C₂₁H₁₅N₃O₃: C, 70.58; H, 4.23; N, 11.76. Found: C, 70.53; H, 4.30; N, 11.73%.

2-(p-Dimethylaminophenyl)-4-hydroxy-4,5-diphenyl-4H-iso*imidazole* (5c). Orange powder, mp 126–128°C; IR (KBr) 1603 (C=N) cm⁻¹; ¹H NMR (500 MHz CDCl₃) δ 3.00 (s, 6H), 6.44 (d, J = 7.5 Hz, 2H), 7.26–7.31 (m, 3H), 7.47 (dd, J = 7.0, 7.53 Hz, 2H), 7.48–7.51 (m, 2H), 7.53 (d, J = 7.5 Hz, 1H), 7.93 (d, J = 7.5 Hz, 2H), 8.35 (d, J = 7.0 Hz, 2H), 6.32 (br s, 1H); MS (FAB) *m/z* 356 (M⁺+1); E.A. Calcd for C₂₃H₂₁N₃O: C, 77.72; H, 5.96; N, 11.82. Found: C, 77.77; H, 5.98; N, 11.81.

2-(*p*-*Hydroxyphenyl*)-4-*hydroxy*-4,5-*diphenyl*-4*H*-*isoimidazole* (5*d*). Yellow powder; mp 103–105°C (dec.); IR (KBr) 3326 (O–H), 1607 (C=N) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.00–7.03 (m, 2H), 7.30–7.34 (m, 5H), 7.41–7.51 (m, 3H), 7.59 (d, J = 7.5 Hz,2H), 8.29 (d, J = 7.5 Hz, 2H). MS (FAB) *m*/*z* 329 (M+H⁺). E.A. Calcd for C₂₁H₁₆N₂O₂·1/ 2H₂O: C, 74.76; H, 5.08; N, 8.30. Found: C, 74.69; H, 5.02; N, 8.81.

2-(*p*-Dimethylaminophenyl)-4-hydroxy-4,5-bis(*p*-fluorophenyl)4H-isoimidazole (5e). Orange powder; mp 138–140°C (dec.); IR (KBr) 1603 (C=N) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 3.04 (s, 6H), 6.64 (d, J = 8.9 Hz, 2H), 6.97 (t, J = 8.8 Hz, 2H), 7.15 (t, J = 9.0 Hz, 2H), 7.40 (d, J = 8.8, 2H), 7.98 (d, J = 8.9 Hz, 2H), 8.35 (d, J = 9.0 Hz, 2H), 6.52 (br s, 1H); E.A. Calcd for C₂₃H₁₉F₂N₃O: C, 67.81; H, 4.70; N, 10.31. Found: C, 67.78; H, 4.76; N, 10.31.

2-(*p*-Dimethylaminophenyl)-4-hydroxy-4,5-bis(*p*-chlorophenyl)4H-isoimidazole (5f). Orange powder; mp 142–144°C (dec.); IR (KBr) 1601 (C=N) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 3.02 (s, 6H), 6.65 (d, J = 8.9 Hz, 2H), 6.90 (t, J = 8.8 Hz, 2H), 7.12 (t, J = 9.0 Hz, 2H), 7.41 (d, J = 8.8, 2H), 7.86 (d, J = 8.9 Hz, 2H), 8.25 (d, J = 9.0 Hz, 2H), 6.49 (br s,





Figure 1. HPLC charts for the resolution.

1H); E.A. Calcd for $C_{23}H_{19}F_2N_3O$: C, 67.81; H, 4.70; N, 10.31. Found: C, 67.35; H, 4.66; N, 10.12.

Mixture of 2-phenyl-4-hydroxy-4-(p-trifluoromethyl-phenyl)-5-(p-fluorophenyl)-4H-isoimidazole (5g) and 2-phenyl-4hydroxy-4-(p-fluorophenyl)-5-(p-trifluoromethyl-phenyl)-4Hisoimidazole (5'g). Molar ratio = 4:1, determined by ¹H NMR. Colorless powder; mp 136–139°C; IR (KBr) 1603 (C=N), 1325 (CF₃), 1272 (C—F) cm⁻¹; UV-vis λ_{max} (CH₂Cl₂) 281 (4.28) nm. MS (FAB) m/z 399 (M⁺+1); E.A. Calcd for C₂₂H₁₄F₄N₂O-1/2H₂O: C, 64.87; H, 3.71; N, 6.88;. Found: C, 64.83; H, 3.75; N, 6.88%. **5g**: ¹H NMR (500 MHz, CDCl₃) δ 7.18–7.28 (m, 4H), 7.38–7.44 (m, 1H), 7.55 (d, J = 8.5 Hz, 2H), 7.61 (d, J = 8.5 Hz, 2H), 7.97 (d, J = 7.5 Hz, 2H), 8.33 (dd, J = 8.5, 5.5 Hz, 2H); **5'g**: ¹H NMR (500 MHz, CDCl₃) 7.03 (t, J = 8.5 Hz, 2H), 7.18–7.27 (m, 2H), 7.37– 7.44 (m, 3H), 7.78 (d, J = 8.2 Hz, 2H), 7.95 (d, J = 7.5 Hz, 2H), 8.46 (d, J = 8.2 Hz, 2H).

Mixture of 2-*phenyl*-4-*hydroxy*-4-(*p*-*trifluoromethyl*-*phenyl*)-5-(*p*-*methoxyphenyl*)-4H-*isoimidazole* (5*h*) and 2-*phenyl*-4-*hydroxy*-4-(*p*-*methoxyphenyl*)-5-(*p*-*trifluoro-methylphenyl*)-4H-*isoimidazole* (5'*h*). Molar ratio = 10:1, determined by ¹H NMR. Pale yellow powder; mp 123–125°C; IR (KBr) 1607 (C=N), 1328 (CF₃), 1263, 1069 (C-O-C) cm⁻¹; UV-vis λ_{max} (CH₂Cl₂) 308 (log ε 4.27), 318 (4.16), 347 (4.12) nm; MS (FAB) *m*/*z* 427 (M⁺+1); E.A. Calcd for C₂₃H₁₇F₃N₂O₂: C, 67.31; H, 4.18; N, 6.83; Found: C, 67.33; H, 4.15; N, 6.85%. **5h:** ¹H NMR (500 MHz, CDCl₃) δ 3.91 (s, 3H), 7.02 (d, *J* = 9.0 Hz, 2H), 7.28 (t, *J* = 7.0 Hz, 2H), 7.37 (t, *J* = 7.0 Hz, 1H), 7.58 (d, *J* = 9.2 Hz, 2H), 7.62 (d, *J* = 9.2 Hz, 2H), 8.01 (d, *J* = 7.0 Hz, 2H), 8.28 (d, *J* = 9.0 Hz, 2H).

Preparation of imidazolones.. 0.5 ml of TBAF/THF (1.0 N) was added to a solution of **5** (0.1 mmol) in DMSO, and the

mixed solution was stirred for 3 h. Then the reaction mixture was poured into water and the product 4 was precipitated. The crude product was purified by chromatography (silica, hexane:AcOEt = 8:1).

2,5,5-Triphenyl-1H-imidazol-4(5H)-one (4a). mp 222–224°C (lit.³ 220–222°C); IR (KBr) 1721 (C=O), 1628 (C=N), 698 (Phenyl) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.28 (t, J = 7.0 Hz, 2H), 7.34 (t, J = 7.0 Hz, 4H), 7.52 (t, J = 7.5 Hz, 2H), 7.58 (t, J = 7.5 Hz, 1H), 7.60 (d, J = 7.0 Hz, 4H), 7.97 (d, J = 7.5 Hz, 2H), 9.11 (br s, 1H); UV-vis λ_{max} (EtOH) 255 (log ε 4.07) nm; MS (FAB) *m*/*z* 313 (M⁺+1); E.A. Calcd for C₂₁H₁₆N₂O: C, 80.75; H, 5.16; N, 8.97. Found: C, 80.47; H, 5.06; N, 8.94%.

2-(*p*-Nitrophenyl)-5,5-diphenyl-1H-imidazol-4(5H)-one (4b.) Pale yellow powder; mp 205–206°C; IR (KBr) 1524 (NO₂), 1350 (NO₂) cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 7.32 (t, 2H), 7,43 (m, 4H), 7.65 (t, 4H), 8.08 (d, J = 9.2 Hz, 2H), 8.18 (d, J = 9.2 Hz, 2H), 9.40 (br s, 1H); MS (FAB) *m*/z 358 (M⁺+1); E.A. Calcd for C₂₁H₁₅N₃O₃: C, 66.75; H, 4.13; N, 11.12. Found: C, 66.73; H, 4.00; N, 11.13%.

2-(*p*-Dimethylaminophenyl)-5,5-diphenyl-1H-imidazol-4(5H)one (4c). Orange powder, mp 86–88°C; IR (KBr) 1604 (C=N) cm⁻¹; ¹H NMR (500 MHz CDCl₃) δ 3.04 (s, 6H), 6.45 (d, J = 7.5 Hz, 2H), 7.26–7.31 (m, 6H), 7.47–7.51 (m, 4H), 7.93 (d, J = 7.5 Hz, 2H), 9.02 (br s, 1H); MS (FAB) *m/z* 356 (M⁺+1); E.A. Calcd for C₂₃H₂₁N₃O: C, 77.72; H, 5.96; N, 11.82. Found: C, 77.70; H, 6.01; N, 11.80.

2-(*p*-*Hydroxyphenyl*)-4,5-*diphenyl*-4*H*-*imidazol*-4(5*H*)-*one* (4*d*). Yellow powder; mp 112–114°C (dec.); IR (KBr) 3323 (O–H), 1605 (C=N) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.05 (d, J = 7.6 Hz, 2H), 7.25–7.31 (m, 6H), 7.45–7.50 (m, 4H), 7.98 (d, J = 7.6 Hz, 2H), 9.02 (br s, 1H). MS (FAB) *m/z* 329 (M+H⁺). E.A. Calcd for C₂₁H₁₆N₂O₂: C, 76.81; H, 4.91; N, 8.53. Found: C, 76.79; H, 4.92; N, 8.51.

2-(*p*-Dimethylaminophenyl)-5,5-bis(*p*-fluorophenyl)-1*H*-imidazol-4(5*H*)-one (4e). Orange powder; mp 118–120°C (dec.); IR (KBr) 1605 (C=N) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 3.06 (s, 6H), 6.67 (d, J = 8.9 Hz, 2H), 6.97–7.15 (t, J = 8.8, 4H), 7.57–7.71 (t, J = 8.8, 4H), 7.92 (d, J = 8.9 Hz, 2H), 9.12 (br s, 1H); MS (FAB) *m*/*z* 392 (M⁺+1); E.A. Calcd for C₂₃H₁₉Cl₂N₃O: C, 65.10; H, 4.51; N, 9.90. Found: C, 65.12; H, 4.46; N, 9.92.

 Table 3

 The conditions of HPLC analysis.

		Retention time/min		ntion /min
	Effluent	wavelength (nm)	Ι	II
5g	Hexane:2-PrOH 80:20 (V/V) ^a	281	6.1	7.1
5h	Hexane:EtOH 90:10 (V/V) ^a	308	6.3	7.8
4g 4h	EtOH 100% ^b EtOH 100% ^b	261 259	5.4 7.0	6.4 9.0

^a Flow rate 1.0 mL/min.

^b Flow rate 0.7 mL/min.



Figure 2. The CD spectra (above) of 5g, h recorded in EtOH and the calculated CD spectra (below) of 5g, h using the TDDFT-B3LYP method. Rotational strengths (*R*) are given in cgs (10^{-40} erg esu cm/Gauss).

2-(*p*-Dimethylaminophenyl)-5,5-bis(*p*-chlorophenyl)-1Himidazol-4(5H)-one (4f). Orange powder; mp 122–124°C (dec.); IR (KBr) 1603 (C=N) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 3.03 (s, 6H), 6.65 (d, J = 8.2 Hz, 2H), 6.90–7.12 (t, J = 8.6 Hz, 4H), 7.61–7.86 (t, J = 8.6, 4H), 7.98 (d, J = 8.2 Hz, 2H), 6.09 (br s, 1H); MS (FAB) *m*/*z* 425 (M⁺+1); E.A. Calcd for C₂₃H₁₉F₂N₃O: C, 67.81; H, 4.70; N, 10.31. Found: C, 67.35; H, 4.66; N, 10.12.

2-Phenyl-5-(p-fluorophenyl)-5-(p-trifluoromethylphenyl)-1Himidazol-4(5H)-one (4g). Colorless crystals; mp 82–88°C; IR (KBr) 1734 (C=O), 1618 (C=N), 1328 (CF₃) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.04 (t, J = 8.8 Hz, 2H), 7.55 (t, J = 7.5Hz, 2H), 7.57–7.64 (m, 5H), 7.75 (d, J = 8.0 Hz, 2H), 7.98 (d, J = 7.5 Hz, 2H), 9.19 (br s, 1H);UV-vis λ_{max} (EtOH) 214 (log ε 4.30), 231 (4.33), 259 (4.02) nm; MS (FAB) m/z 399 (M⁺+1); E.A. Calcd for C₂₂H₁₄F₄N₂O: C, 66.33; H, 3.54; N, 7.03. Found: C, 66.40; H, 3.56; N, 6.98%.

2-Phenyl-5-(p-methoxyphenyl)-5-(p-trifluoromethyl-phenyl)-1H-imidazol-4(5H)-one (4h). Colorless crystals; mp 86–90°C; IR (KBr) 1727 (C=O), 1618 (C=N), 1328 (CF₃), 1253, 1071 (C—O—C) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 3.77 (s, 3H), 6.86 (d, J = 9.0 Hz, 2H), 7.48 (d, J = 9.0 Hz, 2H), 7.53 (t, J = 7.0 Hz, 2H), 7.58 (d, J = 8.3 Hz, 2H), 7.60 (t, J = 7.0 Hz, 1H), 7.75 (d, J = 8.3 Hz, 2H), 8.02 (d, J = 7.0 Hz, 2H); UV-vis λ_{max} (EtOH) 214 (log ϵ 4.33), 232 (4.41) 259 4.08) nm; MS (FAB) m/z 411 (M⁺+1); E.A. Calcd for C₂₃H₁₇F₃N₂O₂: C, 67.31; H, 4.18; N, 6.83; Found: C, 67.40; H, 4.16; N, 6.88%

HPLC analysis. The resolution of the racemic imidazolols **5** and the analysis of the EE were carried out using a chiral HPLC column and a HITACHI 561 recorder, as shown in Figure 1. The conditions are summarized in Table 3.

Assignment of absolute configuration by comparison of experimental and calculated CD spectra. To determine the absolute configuration, the CD spectra were recorded and calculated using the Gaussian 03w software package, [9] as shown in Figures 2 and 3. A geometry optimization was performed using the B3LYP functional with 6-31G* basis sets. The absolute configurations of 5-I and 5-II in the HPLC spectra were assigned as S-5 and R-5, respectively, as shown in



Figure 3. The CD spectra (above) of 4g, h recorded in EtOH and the calculated CD spectra (below) of 4g, h using the TDDFT-B3LYP method. Rotational strengths (R) are given in cgs (10⁻⁴⁰ erg esu cm/Gauss).

Figure 2. In the same manner, the absolute configurations of products **4-I** (HPLC: first fraction) and **4-II** (HPLC: second fraction) were also assigned to **R-4** and **S-4**, respectively, as shown in Figure 3.

REFERENCES AND NOTES

[1] (a) Radziszewski, B. Chem Ber 1877, 10, 70; (b) Dufraisse, C.; Etienne, A.; Martel, J. Comp Rend 1957, 244, 970.

[2] (a) White, E. H.; Harding, M. J. C. J Am Chem Soc 1964,
86, 5686; (b) White, E. H.; Harding, M. J. C. Photochem Photobiol
1965, 4, 1129.

[3] (a) Kimura, M.; Nishikawa, H.; Kura, H.; Lim, H.; White,
E. H. Chem Lett 1993, 505; (b) Kimura, M.; Morioka, M.; Tsunenaga,
M.; Hu, Z. Z. ITE Lett 2000, 1, 418; (c) Hu, Z. Z.; Takami, S.;
Kimura, M.; Tachi, Y.; Naruta, Y. Acta Cryst 2000, C56, e465.

[4] Kimura, M.; Lu, G. H.; Nishigawa, H.; Zhang, Z. Q.; Hu, Z. Z. Luminescence 2007, 22, 72.

[5] (a) Schaap, A. P.; Chen, T. S.; Handley, R. S.; DeSilva, R.; Giri, B. P. Tetrahedron Lett 1987, 28, 1155; (b) McCapra, F. J. Photochem Photobiol A: Chem. 1990, 15, 21; (c) Watanabe, N.; Nagashima, Y.; Yamazaki, T.; Matsumoto, M. Tetrahedron 2003, 59, 4811; (d) Matsumoto, M.; Sakuma, T.; Watanabe, N. Tetrahedron Lett 2002, 43, 8955.

[6] (a) Kimura, M.; Lu, G. H.; Iga, H.; Tsunenaga, M.; Zhang,
 Z. Q.; Hu, Z. Z. Tetrahedron Lett 2007, 48, 3109; (b) Kimura, M.; Lu,
 G. H.; Tsunenaga, M. ITE Lett Batter New Technol Med 2007, 8, 57.

[7] (a) Allen, C. F. H.; VanAllan, J. A. J Am Chem Soc 1943,
65, 1384; (b) Breslow, R.; Chang, H. W.; J Am Chem Soc 1961, 83,
3727; (c) Youssef, A. F.; Ogliaruso, M. J Org Chem 1972, 37, 2601.

[8] (a) Davidson, D.; Weiss, M.; Jelling, M. J Org Chem
1937, 2, 319; (b) Lee, S. H.; Yoshida, K.; Matsushita, H.; Clapham,
B.; Koch, G.; Zimmermann, J.; Janda, K. D. J Org Chem 2004, 69,
8829; (c) Anthony, P.; Adam, D.; Luc, V. H. J Org Chem 2006, 71,
5303; (d) Lovely, C. J.; Du, H.; He, Y.; Dias. R. Org Lett 2004, 6,
735.

[9] Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, J. A., Jr.; Vreven, T.; Kudin, K. N.; Burant, J. C.; Millam, J. M.; Iyengar, S. S.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G. A.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J. E.; Hratchian, H. P.; Cross, J. B.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Ayala, P. Y.; Morokuma, K.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Zakrzewski, V. G.; Dapprich, S.; Daniels, A. D.; Strain, M. C.; Farkas, O.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Ortiz, J. V.; Cui, Q.; Baboul, A. G.; Clifford, S.; Cioslowski, J.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Gonzalez, C.; Pople, J. A. Gaussian 03, Revision B.01; Gaussian, Inc.: Pittsburgh PA, 2003.